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HIPPOCAMPAL PATHOLOGY IN SCHIZOPHRENIA A MORPHOMETRIC STUDY

DISSERTATION

zur Erlangung des Grades eines Doktors der Medizin Der Medizinischen Fakultät der Universität Düsseldorf

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I. INTRODUCTION

Recently, several well controlled morphometric studies of post mortem brains could demonstrate independently pathological alterations in limbic structures of the medial temporal lobe of schizophrenics.

The main findings were altered orientation of hippocampal pyramidal cells (Kovelman and Scheibel, 1984), reduced volumes of the hippocampus, parahippocampal gyrus and amygdala (Bogerts, 1984), enlargement of the inferior horn of the lateral ventricle and reduced thickness of the parahippocampal gyrus (Brown et al., 1986).

These results strongly support the hypothesis that dysfunction of limbic structures of the temporal lobe plays a crucial role in the pathophysiology of schizophrenia (Stevens, 1973, 1982; Torrey and Peterson, 1974).

Furthermore, organic lesions (e.g. tumors, infarctions, traumata) of medial temporal lobe structures are frequently associated with clinical symptoms which are indistinguishable from "endogenous" schizophrenic symptoms suggesting common sites of pathology (Malamud, 1967; Davison and Bagley, 1969; Torrey and Peterson, 1974).

Since pathological alterations within the brain are usually associated with loss of neurons and fibres and increased glial cell densities, tissue volumes and numbers of pyramidal cells and glial cells were determined in all parts of the hippocampal formation, which is the principal structure of the limbic system (Swanson, 1983).

To my knowledge, no such cell counts in the hippocampal formation of schizophrenics have previously been performed.

II. MATERIAL

Complete coronal serial sections of brains from 13 schizophrenic patients (2 male, 11 female; age 43.3 ± 16.8 years (mean \pm SD)

Table 2) and 11 age-matched control cases (7 male, 4 female; 42.6 ± 19.6 , Table 1) of the C. and O. Vogt-Institute of Brain Research, University of Düsseldorf, were investigated.

The time between death and fixation of the brains was about the same in both groups (controls 2-36 h, schizophrenics 4-45 h).

With the exception of two cases - one in each group - only left hemispheres were available for this study.

All brains were collected between 1928 and 1960, fixed by immersion in 4% formalin, embedded in paraffin, cut in 20 µm coronal serial sections and then Nissl-stained and - the adjoining section - myelin-stained. All control cases had no neurological or psychiatric diseases. None of the patients had convulsive, insulin or neuroleptic therapy. The time between first diagnosis of the illness and death, ranged from 10 months to 24 years (mean 9.0 years). 5 of the schizophrenic patients had a predominant paranoid-hallucinatory symptomatology (ICD-9 295.3), 4 schizophrenics had predominant catatonic symptoms (ICD-9 295.2) and 4 had alternating periods of catatonic, paranoid and hebephrenic symptoms (undifferentiated type, mixed type). Further details of patients and controls are given in Table 1 and 2.

III. METHODS

1. Volume determination

All parts of the hippocampal formation were outlined on projected enlargements (10x) of myelin-stained coronal serial sections. Only for volume determination of the granular cell layer Nissl-sections (15x) were used. The distance between the sections was 0.5-1.0 mm, they were evenly distributed from the rostral pole of the hippocampal formation up to the level of the posterior pole of the pulvinar. Details of volume determination by planimetry of serial sections have been described previously (Lesch and Bogerts, 1984).

On myelin-stained sections the following subfields of the hippocampal formation could be clearly delineated (see Figure 1): Alveus and fimbria hippocampi, perforant path, CA1 and CA2 (together), CA3, CA4, prosubiculum and subiculum (together), presubiculum and parasubiculum (together) and the volume of the whole hippocampal formation excluding the parahippocampal gyrus. CA1 and CA2, prosubiculum and subiculum and pre- and parasubiculum were evaluated together, as a reliable separation of these segments was not possible on the myelin-stained sections.

Outlining, planimetry and cell counts were done blindly, i.e. without knowledge of the diagnosis by the individual performing the measurements.

To obtain fresh volumes, the volumes calculated from serial sections were corrected by an average shrinkage factor of 1.89. This value has been determined previously in 30 brains of the Voqt

collection (Lange et al., 1976).

2. Cell densities

Cell densities are expressed as numbers of cells per tissue unit volume.

8 to 10 Niss1 stained sections were evaluated in each brain, 4-5 in the anterior and 4-5 in the posterior portion of the hippocampal formation (Figure 1). The distance between the Niss1 stained sections chosen for cell counts was 1 mm.

In each section, densities of the pyramidal cells were determined by counting nuclei instead of cell bodies at a magnification of x200, densities of glial cells by counting glial nuclei at a magnification of x200. Different types of glial cells were not counted separately. Cell densities were evaluated in the following hippocampal segments: Granular cell layer of the dentate fascia, CA1/CA2, CA3, CA4, subiculum/prosubiculum and presubiculum/parasubiculum. In each section of these areas four microscopic fields were evaluated. Further methodological details of cell counting are given elsewhere (Bogerts, 1977; Bogerts et al., 1983).

3. Absolute cell numbers

Absolute numbers of nerve cells and glial cells were calculated by multiplying numerical cell densities x tissue volume. This value is independent of tissue shrinkage.

IV. CRITERIA OF DELINEATION

1. Myelin stained sections for volume determinations (Figure 1)

The hippocampal formation was defined according to the criteria of Chronister and White (Chronister and White, 1975), including the hippocampus proper, prosubiculum, subiculum, pre- and parasubiculum (Figure 1). The entorhinal region and the whole parahippocampal gyrus (both regions overlap in part) were excluded in this study.

The following criteria were used to separate hippocampal subfields: CA4 situated within the dentate fascia terminates at the hilus of the dentate fascia (Lorente de No, 1934), the mossy fibre system of CA4/CA3 abruptly terminates at the border to CA2 (Braak, 1974). No sharp delineation between CA1 and CA2 was possible, therefore these two segments were evaluated together.

Typical for the prosubiculum and subiculum are myelin bundles crossing the pyramidal cell layer vertically (Rose, 1938; Lorente de No, 1934); the pre- and parasubiculum are characterized by a relative dense plexus of fine myelin fibres (Lorente de No, 1934).

To separate pre- and parasubiculum from the parahippocampal gyrus, an artificial line was drawn using the medial corner of the fissura hippocampi as a topographical marker point as shown in Figure 1; a more precise separation was not possible on myelin sections.

The granular cell layer of the dentate fascia could not clearly be delineated on myelin stained sections; therefore, volume determination and cell counts were performed on the Nissl stained sections.

2. Nissl stained sections for cell counts (Figure 1)

In coronal sections, the structure of the anterior parts of the hippocampal formation differs from that of the posterior parts. CA1, CA2, CA3, and CA4 were delineated according to Rose (Rose, 1934), and Lorente de No (Lorente de No, 1934); the subicular subfields were delineated as described by Braak (Braak, 1972).

V. STATISTICS

Because of the inhomogenous sex distribution, two-way analysis of variance and covariance (diagnosis by sex) with repeated measures on the second factor were performed, using programme package and statistical soft ware of BMDP 7D. Description of groups with histograms and analysis of variance were performed on a Siemens BS 2000 Computer (University of Düsseldorf).

P-values of normals (n) versus (vs) schizophrenics (s) and males (m) vs females (f) are given in the following results.

Mean differences are given in the Tables 3 - 5.

VI. RESULTS

1. Volumes (Table 3)

The volume of the whole hippocampal formation was reduced in the schizophrenic group (n vs s: p = .01), there was no significant sex difference in hippocampal volume.

The volumes of the following hippocampal substructures were significantly reduced in schizophrenics: CA1/CA2 (n vs s: p=.02), CA3 (n vs s: p=.04), CA4 (n vs s: p=.007), pre- and parasubiculum (n vs s: p=.05), granular cell layer of the dentate fascia (n vs s: p=.05).

The whole band of pyramidal cells (CA1 - CA4) was highly significantly reduced (n vs s: p = .003). No volume difference of the subiculum/parasubiculum, alveus and fimbria hippocampi could be found.

The perforant path showed a trend towards volume reduction (n vs p = .10).

2. Absolute numbers of pyramidal cells and glial cells

(Table 4a and 4b)

Schizophrenics exhibit a significant loss of neurons in CA1/CA2 (n vs s: p = .04), CA3 (n vs s: p = .05) and CA4 (n vs s: p = .05).

There was a trend towards a reduction of granular cells of the dentate fascia (n vs s: p = .07).

The cell numbers were unchanged in the prosubiculum/subiculum and

the pre-/parasubiculum.

In contrast to the volume values which revealed no sex differences, the absolute cell numbers of CA3, CA4, prosubiculum/subiculum were lower in females than in males. As shown below, the lower absolute cell numbers in females are due to significant sex dependent differences in cell densities, volumes being unchanged.

3. Glial cells (Table 4b)

Since loss of neurons in brain diseases is usually associated with gliosis, densities and absolute numbers of glial cells were determined without separation in subtypes (i.e. astrocytes, oligodendrocytes, Hortega cells). Surprisingly, the results showed opposite trends for males and females. Schizophrenic males tended to have reduced absolute numbers of glial cells in CA1/CA2 (- 17%), CA3 (- 48%), CA4 (- 28%), prosubiculum/subiculum (- 24%), pre-/parasubiculum (- 51%), schizophrenic females exhibiting increased numbers of glial cells in CA3 (+ 21%), prosubiculum/subiculum (+ 23%), pre-/parasubiculum (+ 9%), in comparison to the controls.

Differences in glial cell numbers were significant for normals versus schizophrenics in CA3 (p = .03), CA4 (p = .05) and pre-/parasubiculum (p = .03).

4. Densities of pyramidal cells (Table 5)

Although there was a loss of pyramidal cells in most segments of the hippocampal formation of schizophrenics, no differences in pyramidal cell densities could be revealed in any of the evaluated areas. Since drop out of nerve cells can cause tissue shrinkage that in turn may lead to an artificial increase in cell densities, absolute cell numbers, which are independent of tissue shrinkage, might be more reliable to assess neuronal loss then cell densities.

Significant sex differences could be detected in CA1/CA2 (n vs s: p = .79), CA3 (n vs s: p = .27), CA4 (n vs s: p = .19) and prosubiculum/subiculum (n vs s: p = .23).

5. Differences between diagnostic subgroups (Figure 2 - 6)

The most distinct loss of hippocampal pyramidal cells occurs in the paranoid-hallucinatory subgroup, in these patients there is a loss of pyramidal cells in CAl/CA2 (n vs s: p = .015, Figure 2), CA3 (n vs s: p = .005, Figure 3), CA4 (n vs s: p = .003, Figure 4), prosubiculum/subiculum (n vs s: p = .001, Figure 5).

The catatonics exhibited a less distinct but also significant loss of pyramidal cells in CA4 (n vs s: p = .03, Figure 4) and dentate fascia (n vs s: p = .03, Figure 6); in the mixed-type subgroup there was a significant reduction of pyramidal cells in CA3 (n vs s: p = .05, Figure 3), CA4 (n vs s: p = .05, Figure 4) and prosubiculum/subiculum (n vs s: p = .02, Figure 5).

Beside the diagnostic classification the following subgroups were

compared:

- patients with hereditary factors versus patients without hereditary factors,
- patients with long duration of illness versus patients with short duration of illness (more or less than 4 years),
- early-onset patients versus young controls (younger than 40 years),
- late-onset patients versus old controls (older than 40 years),
- early-onset cases versus late-onset cases (earlier/later than 40 years).

No significant difference could be found in any of these subgroups.

VII. DISCUSSION

The present data reveal that volume reduction of the hippocampal formation of schizophrenics is associated with reduced numbers of pyramidal cells in the segments CAI/CA2, CA3, CA4, whereas cell numbers of the prosubiculum/subiculum and pre-/parasubiculum are unchanged. There is a trend towards loss of granule cells in the dentate fascia. The reduction of hippocampal pyramidal cells was more distinct in the paranoid-hallucinatory subgroup than in catatonic or undifferentiated schizophrenics.

The major problem in morphometric evaluation of post mortem brain tissue is to ensure that differences between patients and controls reflect actual pathological changes and are not the result of factors unrelated to the disease such as autolysis, duration and kind of the terminal disease, and the histological processing. Three of the controls but only one schizophrenic patient had long lasting terminal diseases (e.g. cancer, Tables 1 and 2), the other controls and schizophrenics had acute causes of death (myocardial infarction, bronchopneumonia, pulmonary embolism). During the long period of collecting the brains (1928 - 1960), the Vogt's and their coworkers took great care of a constant histological processing (fixation, embedding and cutting). The mean interval between death and autopsy was nearly identical in both groups. Thus, it is unlikely that the differences were secondary to a non-cerebral medical illness or due to histological or autolytic artefacts.

Further problems of this study were, that the groups were not sex matched, that numbers of male schizophrenics and female controls were

small, and that nerve cell densities and absolute cell numbers of neurons differed between males and females. Two way analysis of variance and covariance revealed, however, not only significant sex differences but also significant differences between patients and controls. We only investigated brains of patients never treated with neuroleptics, insuline, or electrical shocks; therefore artefacts due to modern medical treatment can be excluded. Nevertheless, we are aware that the results presented are preliminary and need corroboration by studies of larger and sex matched samples. In the Vogt-collection no more schizophrencis and control cases were available.

The values of the mean densities of nerve cells in hippocampal CA-segments of the controls correspond roughly to those found in the human brains by other authors (Ball, 1977; Mouritzen Dam, 1978; Devaney and Johnson, 1984).

Our finding of sexual dimorphism in nerve cell densities is not entirely surprising, since marked sex dependent differences in biochemical and neurophysiological data of the hippocampus have been published previously (Foy et al., 1984; Swaab and Hofmann, 1984; Juraska, 1984).

There exists only one other study which examined quantitatively the histology of the hippocampal formation of schizophrenics (Scheibel and Kovelman, 1984). The authors neither determined volumes of hippocampal subfields nor absolute cell numbers, but found a significant disarray of the pyramidal cells which was interpreted as a morphological correlate of disturbed function of the hippocampus.

Our data, too, indicate that the function of the hippocampus is

altered in schizophrenia, for it is known from all degenerative brain diseases, that reduction of brain tissue and drop out of nerve cells causes impaired functioning of the affected brain parts.

The mean number of glial cells in the schizophrenic group as a whole was not significantly different from controls; therefore reduction of hippocampal nerve cells in schizophrenia is not generally accompanied with gliosis. There were, however, opposite trends for males and females within the schizophrenic group, males exhibiting a decrease, females an increase in glial cell numbers. With respect to the small number of male schizophrenics no definite explanation can be given for this finding, but it is conceivable that at least in a subgroup of schizophrenics, nerve cells and glial cells are affected by the disease.

The present knowledge of the neurophysiology of the hippocampal formation allows an attempt to explain why dysfunction of this structure causes at least some symptoms commonly seen in schizophrenia. Schizophrenics seem to suffer from a lack of "sensory gating" (Asarnow and Mac Crimmon, 1982; Franks et al., 1983; Baribeau-Braun et al., 1983; Schwartz et al., 1983; Siegel et al., 1984), by account of which schizophrenics are unable to filter out non-relevant sensory input and fail to discriminate relevant from non-relevant stimuli. The hippocampus seems to play a crucial role in this "sensory gating", for it receives and integrates informations from all sensory modalities (Papez, 1937; Roberts, 1963; Pandya, 1982; Swanson, 1983). If the information is not familiar, the hippocampus breaks of its inhibitory effect on hypothalamic and brainstem structures involved in attention; the subjects reacts to

the new stimulus. If the stimulus is familiar, it is filtered out; the subject continues with its ongoing behaviour (Pribram and McGiuness, 1975; Schmajuk, 1984). Thus damaged structure of the hippocampus seems to contribute to the complex picture of schizophrenia. Our finding, that the highest loss of neurons occurs in paranoid-hallucinatory patients fits well to reports (Halgreen et al., 1983) that sensory gating is most disturbed in paranoid schizophrenics.

Structural damage of the hippocampus is not specific for schizophrenia, it also occurs in temporal lobe epilepsy (Scheibel et al., 1984; Mouritzen Dam, 1980; Babb et al., 1984) in which CA4 is damaged most, in Alzheimer's disease (Probst et al., 1983; Hyman et al., 1984; Ball, 1985) and Pick's disease (Jellinger and Grisold, 1982). While temporal lobe epileptics sometimes display schizophrenia-like symptoms, Alzheimer and Pick patients do usually not, since only older patients suffer from Alzheimer's disease and Pick's disease and since the schizophrenic symptoms begin usually in adolescence and decline during old age, assume that structural damage of the hippocampus, only under the influence of age related factors (which are not present in senile dementias) predisposes to the development of schiozophrenic symptoms.

In this connection it is of interest that the gonadal steroids testosteron and estrogen have a strong influence on the neuronal activity of hippocampal pyramidal cells (Mc Gowan-Sass, Timiras 1975). Therefore it can be hypothesized that a damaged hippocampus decompensates definitively under the influence of gonadal steroids, thus giving rise to the development of schizophrenic symptoms.

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VIII. REFERENCES

- Asarnow RF, Mac Crimmon DJ (1982) Attention/information processing, neuropsychological functioning, and thought disorder during the acute and partial recovery phases of schizophrenia: a longitudinal study. Psychiatry Res 7: 309-319
- Babb ThL, Lieb JP, Brown JW, Pretorius J, Crandall PH (1984)
 Distribution of pyramidal cell density and hyperexcitability in
 the epileptic human hippocampal formation. Epilepsia 25 (6): 721-728
- Ball MJ (1977) Neuronal loss, neurofibrillary tangles and granulovascuolar degeneration and dementia. Acta Neuropathol, Berlin, 37: 111-118
- Ball MJ, Hachinski V, Fox A, Kirshen AJ, Merskey H, Fisman M, Blume W, Kral VA, Fox H (1985) A new definition of Alzheimer's disease: a hippocampal dementia. The Lancet 1: 14-18
- Baribeau-Braun J, Picton TW, Gosselin JY (1983) Schizophrenia: a neurophysiological evaluation of abnormal information processing. Science 219: 874-876
- Bogerts B (1977) Quantitativ-morphometrische Untersuchungen am Corpus geniculatum laterale und Colliculus superior der Ratte nach vollständigem Lichtentzug. J Hirnforsch 18: 303-319
- Bogerts B, Häntsch J, Herzer M (1983) A morphometric study of the dopamine containing cell groups in the mesencephalon of normals, Parkinson patients and schizophrenics. Biol Psychiatry 18: 951-960
- Bogerts B (1984) Zur Neuropathologie der Schizophrenien. Fortschr Neurol Psychiatr 52: 428-437
- Braak H (1972) Zur Pigmentarchitektonik der Grosshirnrinde des Menschen. II. Subiculum. Z Zellforsch 131: 235-254
- Braak H (1974) On the structure of human archicortex. I. The Cornu Ammonis. Cell Tiss Res 152: 349-383
- Brown et al. (1986) Volume increase of the inferior horn of the lateral ventricle in schizophrenic patients. Arch Gen Psychiatry 43: 36-42
- Chronister RB, White LE (1975) Fiberarchitecture of the hippocampal formation: anatomy, projections, and structural significance. In: Issacson RL, Pribram KH (eds) The Hippocampus, Volume 1: Structure and development. Plenum Press, New York and London, p 21
- Davison K, Bagley CR (1969) Schizophrenia-like psychoses associated with organic disorders of the central nervous system. A review of the literature. In: Current problems in Neuropsychiatry, ed. by

- Hertington RN, Brit J Psychiat Special Publication No. 4, Ashford 1969, pp 113-187
- Devaney KO, Johnson HA (1984) Changes in cell density within the human hippocampal formation as a function of age. Gerontology 30: 100-108
- Foy RF, Chiaia NL, Teyler TJ (1984) Reversal of hippocampal sexual dimorphism by gonadal steroid manipulation. Brain Res 321: 311-314
- Franks RD, Adler LE, Waldo MC, Alpert J, Freeman R (1983) Neurophysiological studies of sensory gating in mania: comparison with schizophrenia. Biol Psychiatry 18 (9): 989-1005
- Halgren E, Wilson CL, Squires NK, Engel J, Walter RD, Crandall PH (1983) Dynamics of the hippocampal contribution to memory: stimulation and recording studies in humans. In: Seifert W (ed) Neurobiology of the Hippocampus. Academic Press, London 1983, pp 2-19
- Hyman BT, Van Hoesen GW, Damasio AR, Barmes CL (1984) Alzheimer's disease: cell-specific pathology isolates the hippocampal formation. Science 225: 1168-1170
- Jellinger K, Grisold W (1980) Zur Morphologie der sogenannten hirnatrophischen Prozesse. In: Das ärztliche Gespräch 34 "Neuropathologisches Symposium" Haar 1980, Frydl V (ed) Herausgegeben von der Medizinischen Abteilung der Troponwerke Köln
- Juraska JM (1984) Sex differences in developmental plasticity in the visual cortex and hippocampal dentate gyrus. In: De Vries GJ et al. (eds) Progress in Brain Research Vol 61, Elsevier Science Publishers B.V., Amsterdam, pp 205-214
- Lange H, Thörner G, Hopf A, Schröder KF (1976) Morphometric studies of the neuropathological changes in choreatic diseases. J Neurol Sci 28: 401-425
- Lesch A, Bogerts B (1984) The diencephalon in schizophrenia: evidence for reduced thickness of the periventricular grey matter. Eur Arch Psychiat Neurol Sci 234: 212-219
- Lorente de No R (1934) Studies on the structure of the cerebral cortex. II. Continuation of the study of the ammonic system. J Psychol Neurol 46 (2, 3): 113-177
- Malamud N (1967) Psychiatric disorder with intracranial tumors of the limbic system. Arch Neurol 17: 113-123
- Mc Gowan-Sass BK, Timiras PS (1975) The hippocampus and hormonal cyclicity. In: Isaacson RL, Pribram KH (eds) The Hippocampus. Vol. 2: Neurophysiology and behavior. Plenum Press, New York and London, pp 355-370
- Mouritzen Dam A (1979) The density of neurons in the human hippocampus. Neuropathol Appl Neurobiol 5: 249-264

- Pandya DH, Seltzer B (1982) Association areas of the cerebral cortex. Trends Neurosci 5: 386-390
- Papez JW (1937) A proposed mechanism of emotion. Arch Neurol Psychiatr 38: 725-743
- Pribram KH, Mc Guiness (1975) Summary. In: Isaacson RL, Pribram KH (eds) The Hippocampus. Vol 2: Neurophysiology and behavior. Plenum Press, New York and London, pp 429-441
- Probst A, Basler V, Bron B, Ulrich J (1983) Neuritic plaques in senile dementia of Alzheimer type: a Golgi analysis in the hippocampal region. Brain Res 268: 249-254
- Roberts DR (1963) Schizophrenia and the brain. J Neuropsychiatr 5: 71-19
- Rose J (1938) Zur normalen und pathologischen Architektur der Ammonsformation. J Psychol Neurol 49 (1, 2): 137-186
- Scheibel AB, Kovelman JA (1984) A neurohistological correlate of schizophrenia. Biol Psychiatry 19 (12): 1601-1621
- Scheibel ME, Crandall PH, Scheibel AB (1974) Complex in temporal lobe epilepsy. Epilepsia 15: 55-80
- Schmajuk NA (1984) Psychological theories of hippocampal function. Physiol Psychol 12 (3): 166-183
- Schwartz BD, Winstead DK, Adinoff B (1983) Temporal integration deficit in visual information processing by chronic schizophrenics. Biol Psychiatry 18 (11): 1311-1320
- Siegel C, Waldo M, Mizner G, Adler LE, Freedman R (1984) Deficits in sensory gating in schizophrenic patients and their relatives. Arch Gen Psychiatry 41: 607-612
- Stevens JR (1973) An anatomy of schizophrenia? Arch Gen Psychiatry 29: 177-189
- Stevens JR (1982) Neuropathology of schizophrenia. Arch Gen Psychiatry 39: 1131-1139
- Swaab DF, Hofman MA (1984) Sexual differentiation of the human brain. De Vries et al. (eds) Progress in Brain Research, Vol 61, Elsevier Science Publishers, Amsterdam, pp 361-373
- Swanson LW (1983) The hippocampus and the concept of the limbic system. In: Seifert W (ed) Neurobiology of the Hippocampus. Academic Press, London 1983, pp 2-19
- Torrey EF, Peterson MR (1974) Schizophrenia and the limbic system. Lancet 2: 942-946

IX. TABLES

В	rain	Age	Sex	Cause of death				
A	56	56	m	Laryngeal cancer, death under operation				
A	58	24	m	Haemorrhagic shock				
A	61	38	m	Uraemic coma				
A	64	84	m	Pneumonia				
A	80	33	f	Carcinoma of the uterus				
A	81	29	f	unknown				
A	85	30	f	Fat embolism after a street accident				
A	88	62	m	Carcinoma of the stomack, peritonitis				
A	97	39	m	Pulmonary embolism				
A	100	19	m	Aspiration pneumonia				
A	102	41	f	Cardiac arrest				

 $\underline{\text{Table 1}}$: Control cases without neurological or psychiatric disease. Brains were collected between the years 1928 and 1960.

Brain	Age	Sex	Cause of death year of death	Most prominent psychiatric symptoms	Time between diagnosis and death		
Bu 3	22	f	Cardiac arrest 1930	Persecution complex, acoustic halluci- nations, mutism, negativism	3 1/2	years	
Bu 7	74	f	Myocardial in- farction, 1930	Paranoid ideas, acoustic hallucina- tions, mutism, manierism	24	years	
Bu 9	59	f	Bronchopneumonia 1930	Paranoid ideas, optic hallucinations, coenaesthetic symptoms	4	years	
Bu 12	51	f	Bronchopneumonia 1930	Persecution complex, stupor, negativism, mutism	not	known	
Bu 19	44	f	Pneumonia 1930	Paranoid ideas, feeling of being watched, acoustic hallucinations	11	months	
Bu 20	26	f	Bronchopneumonia 1930	Stupor, catatonia, mutism	7	years	
Bu 21	42	f	Pneumonia 1930	Stupor, mutism, flexibilitas cerea, acoustic hallucinations	6 1/2	years	
Bu 24	27	f	Bronchopneumonia 1930	Optic, acoustic and haptic halluci- nations, stereotype movements, manierism, hebephrenic symptoms	4	years	
Bu 46	19	f	Cardiac illness 1932	Rigid postures, drawing faces, negativism	not	known	
Bu 52	24	m	Bronchopneumonia 1933	Stupor, stereotypy in posture and movement, catatonia, catalepsy	3	years	
Bu 62	64	f	Suicide 1933	Delusions, hallucinations, persecution complex	23	years	
Bu 89	43	f	Pneumoria 1935	Acoustic hallucinations, paranoid ideas, feeling of electrical stimulation	13	months	
CP 81	39	m	not known 1941	Hebephrenic symptoms, drawing faces, delusions, manierism, mutism	22	years	

<u>Table 2</u>: Schizophrenic cases. The brains were collected before the introduction of neuroleptic drugs, between the years 1930 and 1941.

Evaluated areas	Controls		Schizophrenics		mean diff. (Controls = 100 %)			p-values	
	Males (n=7)	Females (n=4)	Males (n=2)	Females (n=11)	m	f	m	vs f	c vs s
	Mean (SD)	Mean (SD)	Mean (SD)	Mean (SD)	(%)	(%)			
total hippocampal	3493	3354	2708	2942	-22	-12	Р	= .83	p = .01
formation CA1/CA2	(427) 431 (67)	(403) 574 (83)	(886) 393 (59)	(384) 404 (95)	- 9	-30	р	= .08	p = .02
CA3	826 (145)	706 (132)	535 (116)	680 (158)	-35	+ 4	P	= .87	p = .04
CA4	445 (52)	459 (101)	339 (124)	343 (69)	-24	-25	P	= .81	p = .00
total pyramida band of the hippo- campal formation	2200 (244)	2190 (239)	1638 (376)	1861 (275)	-26	-14	Р	= .43	p = .00
prosubiculum/ subiculum	227 (53)	197 (36)	236 (74)	194 (45)	+ 4	- 2	p	= .15	p = .91
presubiculum/ parasubiculum	280 (77)	270 (68)	180 (66)	237 (56)	-29	-12	P	= .48	p = .05
granular cell laye of dentate fascia	(189)	1026 (233)	774 (439)	709 (176)	-12	-31	P	= .68	p = .05
alveus and fimbria hippocampi	(115)	444 (81)	393 (150)	428 (64)	-24	- 4		= .68	p = .13
perforant path	775 (9 6)	682 (137)	603 (249)	654 (97)	-22	- 4	Р	= .71	p = .10

<u>Table 3</u>: Tissue volumes of the hippocampal formation and all hippocampal segments (values in mm³).
Mean, standard deviation (SD), number of sample (n), mean differences given separately for male (m) and female (f) cases. p-values of controls (c) versus (vs) schizophrenics (s) and males (m) versus females (f).

Evaluated areas	Controls		Schizophrenics		mean diff. (Controls = 100 %)	p-values	p-values	
	Males (n=7)	Females (n=4)	Males (n=2)	Females (n=11)	m f	m vs f c vs	s	
	Mean (SD)	Mean (SD)	Mean (SD)	Mean (SD)	(%) (%)			
CA1/CA2	27 (4)	32 (9)	25 (1)	21 (6)	- 6 -36	p = .88 p =	.04	
CA3	54 (11)	37 (11)	39 (3)	34 (7)	-28 - 7	p = .03 p =	.05	
CA4	21 (3)	13 (3)	15 (3)	12 (4)	-28 - 4	p = .003 p =	.05	
prosubiculum/ subiculum	10 (2)	7 (1)	11 (2)	8 (2)	+14 +15	p = .004 p =	. 20	
presubiculum/ parasubiculum	8 (3)	8 (2)	7 (.5)	7 (2)	-19 -19	p = .85 p =	.17	
granular cell laye	er 18 (6)	23 (8)	14 (11)	15 (4)	-20 -32	p = .31 p =	.07	

 $\underline{\text{Table 4a}}$: Absolute numbers of pyramidal cells (x 10^5). For abbreviations see Table 3.

Evaluated areas	Controls		Schizophrenics		mean diff. (Controls = 100 %)		p-values		
	Males (n=7)	Females (n=4)	n=4) Males (n=2)	Females (n=11)	m	f	m vs f	c vs s	
• • • • • • • • • • • • • • • • • • • •	Mean (SD)	Mean (SD)	Mean (SD)	Mean (SD)	(%)	(%)			
CA1/CA2	96 (16)	102 (29)	80 (4)	90 (20)	-17	-11	p = .41	p = .17	
CA3	270 (60)	169 (21)	139 (25)	204 (31)	-48	+21	p = .38	p = .03	
CA4	142 (24)	128 (35)	101 (41)	110 (26)	-28	-13	p = .83	p = .05	
prosubiculum/ subiculum	94 (20)	66 (5)	71 (6)	80 (19)	-24	+23	p = .30	p = .63	
presubiculum/ parasubiculum	148 (38)	113 (21)	73 (18)	122 (21)	-51	+ 9	p = .62	p = .03	

 $\frac{\text{Table 4b}}{\text{For abbreviations see Table 3.}}: \text{Absolute numbers of glial cells (x } 10^5).$

Evaluated areas	Controls		Schizophrenics		mean diff. (Controls = 100 %)		p-values		
	Males (n=7)	Females (n=4)	Males (n=2)	Females (n=11)	m	f	m vs f	c vs s	
	Mean (SD)	Mean (SD)	Mean (SD)	Mean (SD)	(%)	(%)			
CA1/CA2	63 (7)	49 (6)	59 (11)	51 (11)	- 6	+ 4	p = .03	p = .79	
CA3	66 (7)	50 (5)	75 (20)	52 (11)	+14	+ 4	p = .001	p = .27	
CA4	47 (7)	31 (6)	50 (6)	39 (9)	+ 7	+24	p = .002	p = .19	
prosubiculum/ subiculum	44 (5)	34 (4)	45 (7)	41 (8)	+ 2	+22	p = .05	p = .23	
presubiculum/ parasubiculum	31 (8)	31 (2)	35 (11)	32 (5)	+13	+ 3	p = .53	p = .43	
granular cell laye	er 204 (39)	198 (19)	210 (60)	222 (44)	+ 3	+12	p = .87	p = .49	

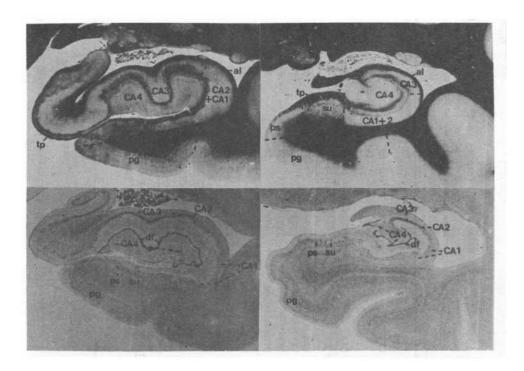
 $\frac{\text{Table 5}}{\text{For abbreviations see Table 3.}}: \text{ Densities of pyramidal cells (x 100/mm}^3).}$

X. FIGURE LEGENDS

Figure 1

Two myelin-(top) and two Nissl-stained (bottom) coronal sections of the anterior part (left) and the posterior part (right) of the hippocampal formation (magnification 6x) showing the different anatomy of the CA segments and of the dentate gyrus at the two levels. Distance between the sections is about 2 cm. Criteria of delineation are given in the text.

al = alveus, tp = tractus perforans, df = dentate fascia, CAl - CA4 = hippocampus proper, su = prosubiculum/subiculum, ps = presubiculum/parasubiculum, pg = parahippocampal gyrus



Figures 2 - 6:

Absolute numbers of nerve cells of controls (c, first column), catatonic (cat, second column), paranoid-hallucinatory (par, third column), and mixed-type (mixed) schizophrenics. Mean, number of cases, difference to controls in %, p-value (normals versus schizophrenics, F-test), ns = not significant.

Figure 2: Hippocampal Segments CA 1 + CA 2

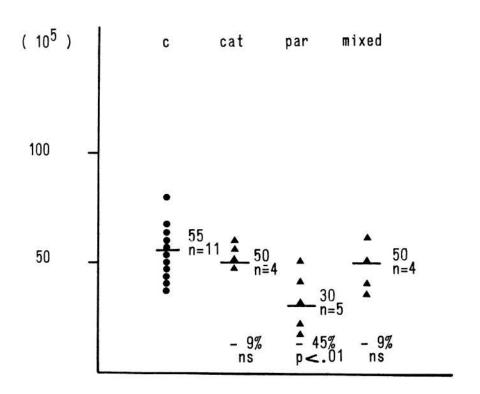


Figure 3: Hippocampal Segment CA 3

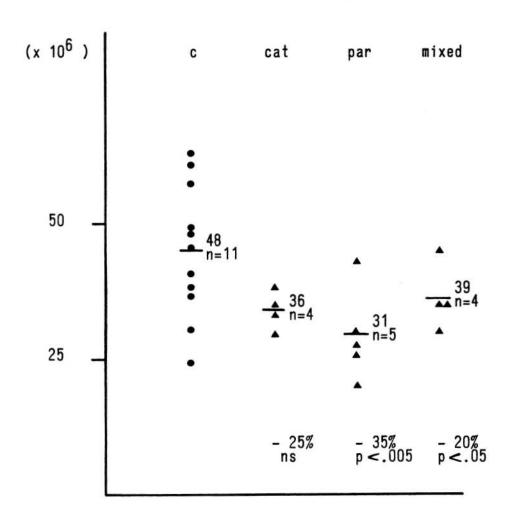


Figure 4: Hippocampal Segment CA 4

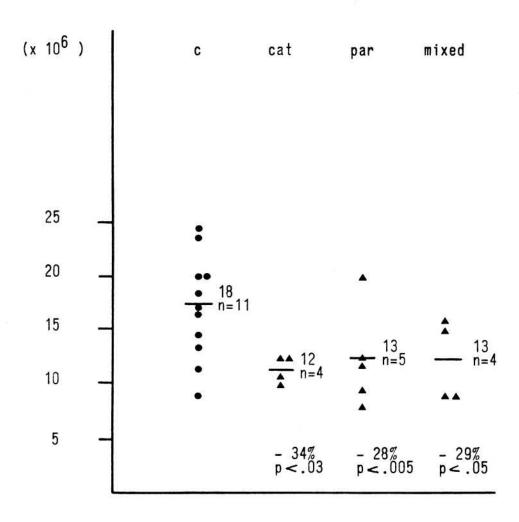
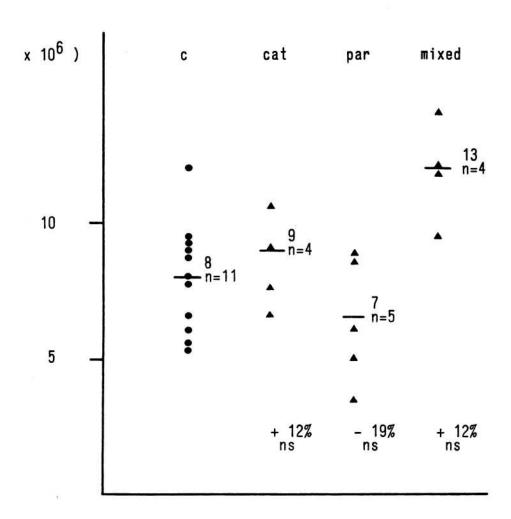
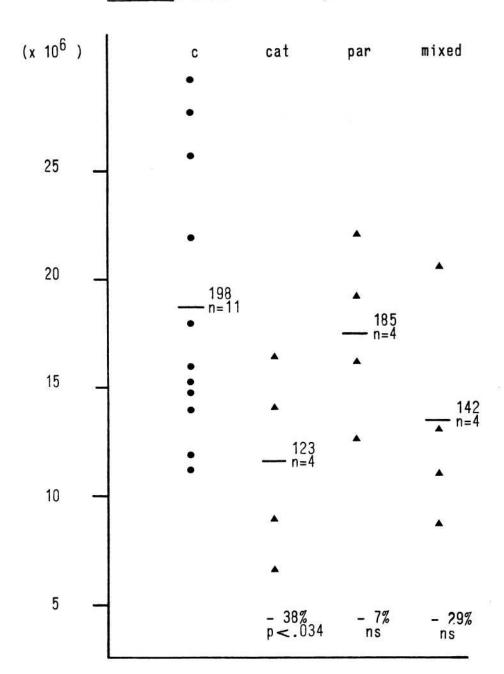


Figure 5: Subiculum





XI. SUMMARY

To investigate whether volume reduction of the hippocampal formation of schizophrenics, as described previously, is paralleled by loss of neurons and fibre systems, tissue volumes and cell numbers of all parts of the hippocampal formation in post mortem brains of 13 schizophrenics and 11 age-matched controls belonging to the Vogt-collection were determined.

Volumes of the whole hippocampal formation (p = .01), the whole pyramidal band (p = .001) and the hippocampal segments CAl/CA2 (p = .01), CA3 (p = .05), CA4 (p = .01) were decreased, where as no significant volume reduction of the alveus and fimbria hippocampi and prosubiculum/subiculum could be found. The perforant path showed a trend towards volume reduction (p = .1).

The absolute number of pyramidal cells (tissue volume \times cell density) was diminished in CAl/CA2 (p = .05), CA3 (p = .05) and CA4 (p = .05), but was not significantly changed in the prosubiculum/subiculum, the presubiculum/parasubiculum and the granular cell layer of the dentate fascia.

Pyramidal cell loss in CA1/CA2, CA3, CA4 was more distinct in the paranoid patients than in catatonics.

The findings are discussed with respect to current hypotheses of limbic dysfunction in schizophrenia.

Key words : schizophrenia, hippocampus, neuropathology